

The single CCA-adding enzyme of *T. brucei* has distinct functions in the cytosol and in mitochondria

Shikha Shikha¹ and André Schneider^{1*}

Running title: CCA-adding enzyme of *T. brucei*

From the ¹Department of Chemistry and Biochemistry, University of Bern, Freiestrasse 3, Bern CH-3012, Switzerland

* To whom correspondence should be addressed: André Schneider: Department of Chemistry and Biochemistry, University of Bern, Freiestrasse 3, Bern CH-3012, Switzerland; andre.schneider@dcb.unibe.ch; +41 31 631 4253

Keywords: tRNA, CCA-adding enzyme, mitochondria, trypanosomes,

ABSTRACT

tRNAs universally carry a CCA nucleotide triplet at their 3'-ends. In eukaryotes, the CCA is added post-transcriptionally by the CCA-adding enzyme (CAE). The mitochondrion of the parasitic protozoan *Trypanosoma brucei* lacks tRNA genes and therefore imports all of its tRNAs from the cytosol. This has generated interest in the tRNA modifications and their distribution in this organism, including how CCA is added to tRNAs. Here, using a BLAST search for genes encoding putative CAE proteins in *T. brucei*, we identified a single ORF, Tb927.9.8780, as a potential candidate. Knockdown of this putative protein, termed TbCAE, resulted in the accumulation of truncated tRNAs, abolished translation, and inhibited both total and mitochondrial CCA-adding activities, indicating that TbCAE is located both in the cytosol and mitochondrion. However, mitochondrially localized tRNAs were much less affected by the TbCAE ablation than the other tRNAs. Complementation assays revealed that the N-terminal 10 amino acids of TbCAE are dispensable for its activity and mitochondrial localization and that deletion of 10 further amino acids abolishes both. A growth arrest caused by the TbCAE

knockdown was rescued by the expression of the cytosolic isoform of yeast CAE, even though it was not imported into mitochondria. This finding indicated that the yeast enzyme complements the essential function of TbCAE by adding CCA to the primary tRNA transcripts. Of note, ablation of the mitochondrial TbCAE activity, which likely has a repair function, only marginally affected growth.

Unlike in most other eukaryotes the mitochondrion of the parasitic protozoan *T. brucei* and its relatives does not encode any tRNAs (1,2). However, organellar tRNAs are required for mitochondrial translation which is essential in both the insect-stage and the bloodstream form of the parasite (3,4). Thus, all mitochondrial tRNAs must be imported from the cytosol. Mitochondrial tRNA import is not restricted to trypanosomes but has been found in many organisms including some fungi, plants and protozoans (2,5-7). But in essentially all other systems, at least a subset of tRNAs is still encoded in the mitochondrial genome. tRNA import in trypanosomes has been studied in quite some detail and while there are still many unanswered questions regarding the process, some factors mediating tRNA import have

recently been characterized (8-10). As in other systems a connection to mitochondrial protein import has emerged, even though in contrast to what has been proposed in yeast, trypanosomal tRNAs are not co-imported with proteins (9). Moreover, tRNAs appear to be imported as fully processed molecules (11,12). The exclusive reliance on mitochondrial tRNA import has initiated a more general interest in trypanosomal tRNA biology (1). Aminoacyl-tRNA synthetases (aaRSs) have been studied extensively and it was shown that, likely due to mitochondrial tRNA import, most but interestingly not all aaRSs are dually targeted to the cytosol and the mitochondrion (3,13-15). Furthermore, a mitochondrion-specific editing of the anticodon of the tRNA^{Trp} has been shown, that allows it to decode different codons in the cytosol and the mitochondrion, explaining how *T. brucei* accommodates the re-assignment of the stop codon UGA to Trp in mitochondria (16). This has fostered extensive work on tRNA modifications in general (17-19). Trypanosomes were also instrumental in deciphering the eukaryotic pathway for selenocysteinylation of tRNA^{Sec} formation (20). Studies on that subject were aided by the fact that selenoproteins are neither essential for procyclic nor bloodstream forms of the parasite when cultured in vitro (21). Furthermore, unlike in other systems and in contrast to all other trypanosomal tRNAs, the tRNA^{Sec} of *T. brucei* is transcribed by RNA Polymerase II as it is embedded in a polycistron (22).

Eukaryotic tRNAs are transcribed by RNA polymerase III as precursors carrying 5'- and 3'-extensions and may carry introns. Correct processing of the precursor is essential for aminoacylation and thus for tRNA function (23,24). 5'-processing of tRNAs is generally catalyzed by RNase P, which in eukaryotes consists of a variable number of protein factors and of an RNA subunit, that carries the enzymatic activity. In trypanosomes this is different, a recent study has shown that *T. brucei* has an unusual protein-only RNase P and a similar situation has also been observed in plants (25,26). Furthermore, a mitochondrial version of a protein-only RNase P enzyme has also been characterized in *T. brucei*, even though it is not clear what the in vivo substrate of that enzyme would be (26,27). 3'-processing of tRNAs involves cleavage of the 3'-extensions by RNases including RNase Z

(24,28). Subsequently CCA is added to the processed 3'-end by the non-templated CCA-adding enzyme (CAE) creating the universal 3' CCA end that is found on all tRNAs (while in many bacteria the CCA is already encoded in the tRNA gene this not the case in eukaryotes) (29). It is the 3'-terminal A that gets aminoacylated by the aaRSs. 3'-processing of tRNAs is one of the few aspects of tRNA biology that has so far not been studied in trypanosomes. This was the reason why we decided to characterize the trypanosomal CAE. In the present study, we show that the enzyme is essential for normal growth, because its ablation abolishes translation. We furthermore show that the protein is dually localized in mitochondria and the rest of the cell.

RESULTS

TbCAE is essential for cytosolic translation

Doing BLAST searches using the sequences of the yeast or human CAEs as templates we identified a single ORF, Tb927.9.8780, that appears to encode the trypanosomal CAE, which we termed TbCAE. In order to functionally study the enzyme, we produced an inducible RNAi cell line in procyclic *T. brucei* that targets the ORF of the mRNA. Fig. 1A shows that addition of tetracycline results in the efficient ablation of the TbCAE mRNA. Moreover, growth slows down after one day and stops after two days RNAi induction, which is typical for an essential gene.

Ablation of the CAE should ultimately abolish translation as it would lead to accumulation of non-chargeable tRNAs with immature 3'-ends. In order to test this, we labeled cell culture with ³⁵S-methionine which gets incorporated into proteins during translation (Fig. 2B, left two lanes). When the experiment was done with the induced TbCAE-RNAi cell line we observed a great reduction of ³⁵S-methionine incorporation after 24 hours of induction when compared to uninduced cells. Moreover, after 48 hours translation was essentially abolished. Virtually the same was seen if cells were treated with the translation inhibitor cycloheximide. Thus, ablation of TbCAE, as might be expected, rapidly abolishes translation.

Ablation of TbCAE inhibits its activity

In order to directly measure TbCAE activity we prepared total cellular extracts of the uninduced

and 2 day induced TbCAE-RNAi cell line. The extracts were incubated in the presence of radioactive α - 32 P-CTP and isolated total RNA (30). Subsequently, total RNA was re-isolated and separated by 8M Urea/15% polyacrylamide-electrophoresis (PAGE). The gel was stained by ethidium bromide, dried and exposed on a Phosphorimager. The result in the upper left panel of Fig. 2 shows a highly efficient labeling of the tRNA region that was greatly reduced after ablation of TbCAE. The same result was obtained when α - 32 P-ATP but not when α - 32 P-UTP was added to the reaction. We concluded from these results that C and/or A were added to the small fraction of the newly synthesized and 3'-cleaved tRNAs present in the assay mixture. Thus, our results strongly suggest that TbCAE adds both C and A to the 3'-ends of newly transcribed and processed trypanosomal tRNAs. The absence of labeling in the presence α - 32 P-UTP indicates that it is not caused by transcription.

Ablation of TbCAE differentially affects different tRNAs

In order to investigate how ablation of TbCAE affects different tRNA isoacceptor groups in vivo we isolated total RNA from uninduced and two days induced cells, separated it by 8M Urea/15% PAGE and stained the gel by ethidium bromide (Fig. 3, lower panels). The tRNA regions of total RNA isolated from uninduced and induced cells look very similar. In order to look at individual tRNAs we analyzed the steady-state levels of all tRNA isoacceptor groups by Northern blotting using specific oligonucleotide probes (Fig. 3, upper panels). The results allow the separation of the tRNAs into four groups regarding how they react to the ablation of TbCAE.

- The strongest phenotypes were seen for tRNA^{Cys}, tRNA^{His} and for the tRNA-like molecule that is part of the trypanosomal signal recognition particle (31). In all three cases we see efficient accumulation of a lower molecular weight band, suggesting that tRNAs lacking a part or the whole CCA-tail accumulate. In the case of tRNA^{Cys} and tRNA^{His} the truncated molecules are the main species observed.

- For tRNA^{Arg}, tRNA^{Gln}, tRNA^{Trp}, we see the same as above except that only very small amounts of truncated tRNAs are found.

- For tRNA^{Ala}, tRNA^{Gly} and tRNA^{Thr}, ablation of TbCAE causes a significant reduction in the

steady-state levels when compared to the total tRNAs seen in the ethidium bromide stained gel, that serves as a loading control. However, no specific accumulation of lower molecular weight forms is seen.

- Finally, and perhaps most surprisingly we find that 14 tRNA acceptor groups are not affected by the knock down of the TbCAE.

For some tRNAs (Fig. 3, tRNA^{Met}) we see additional low molecular weight bands. These might be other tRNA isoacceptor that crossreact with the oligonucleotide that was used as a probe. Alternatively they may represent hypo-modified tRNAs in the cases where the modification was added after the CCA addition.

tRNA are known to be quite stable molecules. Thus, a caveat of our analysis is that the tRNA population, that was synthesized before the RNAi takes effect, might not be affected. In order to increase the sensitivity of the analysis we used a cell line in which upon addition of tetracycline not only TbCAE-RNAi is induced but also expression of a mutationally tagged tRNA^{Met} occurs (11). (This is possible because its gene contains a 5'-flanking tetracycline-operator sequence). With this system, we can selectively monitor the fate of the newly synthesized variant of tRNA^{Met} (tRNA^{Met*}) during induction of TbCAE RNAi. Interestingly, a significant accumulation of truncated tRNA^{Met*} species was seen in such an experiment (Fig. 4), even though no phenotype was observed for either of the two endogenous tRNAs^{Met} (tRNA^{Met-i} and tRNA^{Met-e}) when their steady state levels were analyzed (Fig. 3).

In summary, these results show that ablation of TbCAE leads to accumulation of truncated tRNAs as would be expected. However, surprisingly there were large differences how much truncated tRNAs accumulated, from 72.02% for tRNA^{Cys} (SD \pm 2.92%) to a very low percentage for tRNA^{Met}, that could only be detected when newly synthesized tRNA was analyzed. RNAi inherently does not completely remove its target mRNA, indicating that a small amount of TbCAE is still present in induced cells. Assuming a similar half-life for all tRNA isoacceptor groups, the differential effects we see might be explained by different affinities of TbCAE to different tRNA isoacceptor groups. Thus, the tRNAs that are only marginally affected may have higher affinities for the enzyme and therefore the CCA-tail might still

be added to these substrates even when only very little enzyme is present in induced RNAi cells.

The simplest way to explain the reduced levels of tRNA^{Ala}, tRNA^{Gly} and tRNA^{Thr} observed in induced RNAi cells is that the truncated tRNA lacking the CCA-tail get rapidly degraded.

A fraction of TbCAE is imported into mitochondria

In order to determine the intracellular localization of TbCAE, we produced a transgenic cell line expressing a version of TbCAE whose C-terminus is tagged with the c-myc epitope. Fig. 5A shows that the TbCAE-myc is expressed. Moreover, when the cells are extracted with low concentration of digitonin the protein is essentially recovered in the soluble supernatant fraction. In the pellet which contains intact mitochondria approximately the same percentage of total tagged TbCAE is detected as is for the cytosolic markers elongation factor 1a (EF1a) and TbVCP. In line with this, TbCAE-myc is localized all over the cell when analyzed by immunofluorescence (Fig. 5B). In summary these results suggest that the large majority of TbCAE is cytosolically and to a lesser extent nuclearly localized. In other eukaryotic systems a fraction of CAEs is generally imported into mitochondria (32-35). The experiments shown in Fig. 5AB cannot confirm this for *T. brucei*. Moreover, the N-terminus of TbCAE is not predicted to form a mitochondrial targeting sequence. However, not all mitochondrial proteins have typical N-terminal targeting sequences and import of a small fraction of TbCAE might be beyond the detection limit in these experiments. In fact, in a recent study we have experimentally defined the mitochondrial proteome of *T. brucei* using two criteria (15): to be scored as a component of the mitochondrial proteome a protein needs to be i) present in a highly enriched gradient-purified mitochondrial fraction and ii) its levels have to be reduced in mitochondria ablated for ATOM40, the import pore of the main translocase of the outer membrane (36,37). TbCAE fulfilled both criteria in this study and therefore is a bona fide component of the mitochondrial proteome (Fig. 5C) (15).

Should a small fraction of TbCAE be mitochondrially localized we would expect that its activity can be detected in mitochondrial extracts using the same assay that was used for

total RNA (Fig. 2). Fig. 6 shows that this is the case. Mitochondrial tRNAs are strongly labeled with α -³²P-CTP in an organellar extract, however after depletion of TbCAE by RNAi the labeled fraction is dramatically reduced. The immunoblots in Fig. 6B confirm that the mitochondrial extracts used in activity assays were not contaminated by cytosolic proteins.

These results confirm that TbCAE is indeed a component of the mitochondrial proteome as has recently been determined (15), even though standard biochemical and cell biological methods are not sensitive enough to detect this (Fig. 5AB).

Ablation of TbCAE only moderately affects mitochondrial tRNAs

As was done for total RNA (Fig. 3) we analyzed the effect ablation of the mitochondrial TbCAE activity has on the steady state level of the different tRNA isoacceptor groups that were imported into mitochondria. To that end organellar RNA was isolated from digitonin extracted uninduced and induced TbCAE RNAi cells that were treated with RNase to digest the contaminating cytosolic RNAs. Fig. S2 shows that the resulting mitochondrial RNA fractions, in contrast to total cellular RNA, is free of the cytosol-specific initiator tRNA^{Met} and tRNA^{Sec}.

In contrast to when tRNAs from total cellular extracts (of which less than 5% correspond to mitochondrial tRNA (38)) were analyzed, mitochondrially imported tRNAs do not seem to be affected by ablation of TbCAE (Fig. 7). The one exception is tRNA^{Cys} for which accumulation of a small fraction of a lower molecular weight form is observed. Interestingly, tRNA^{Cys} also shows the strongest phenotype when total tRNAs were analyzed (Fig. 3). For tRNA^{Gln} and tRNA^{His} we detected higher steady state levels after ablation of TbCAE.

To increase the sensitivity of our assay we again used the cell line in which upon addition of tetracycline both ablation of TbCAE and expression of the mutationally tagged tRNA^{Met} are induced. However, instead of analyzing total tRNA (Fig. 4) we analyzed the mitochondrial tRNAs that were free of cytosolic tRNA (Fig. S3). The results show that ablation of TbCAE causes an approximately 50% accumulation of a truncated newly synthesized mitochondrially-localized tRNA^{Met} variant (Fig. 8).

In summary, these results indicate that ablation of TbCAE affects at least a subset of mitochondrially imported tRNAs. However, the extent to which truncated forms accumulate is much lower than for total tRNA.

Complementation experiments using truncated TbCAE variants

To perform *in vivo* complementation experiments we used a synthetic gene that encodes a C-terminally myc-tagged version of the TbCAE that is resistant to RNAi due to multiple synonymous codon replacements. Expression of this synthetic gene in the induced TbCAE-RNAi cell line resulted in essentially wildtype growth (Fig. 9A). This confirms that i) the phenotypes observed in the TbCAE-RNAi cell line described above are not caused by off target effects and that ii) the C-terminal myc-tag does not interfere with the function of the enzyme. Next, we transfected the TbCAE-RNAi cell line with constructs allowing inducible expression of three N-terminal truncated variants of TbCAE lacking either 10, 20 or 30 N-terminal amino acids, termed ΔN_{10} TbCAE-myc, ΔN_{20} TbCAE-myc and ΔN_{30} TbCAE-myc, respectively (Fig. 9B-D). All complemented cell lines were tested for ablation of the endogenous TbCAE mRNA, growth, expression and localization of the myc-tagged TbCAE variants as well as for TbCAE activity. The results show that in all cell lines the endogenous TbCAE mRNA was downregulated and that the transgenic TbCAE variants were expressed (Fig. 9, insets in growth curve panels). Moreover, while the cell line expressing ΔN_{10} TbCAE-myc grew normal (Fig. 9B), like the cell line complemented with the full-length CAE (Fig. 9A), expression of the ΔN_{20} TbCAE-myc and ΔN_{30} TbCAE-myc variants did not restore growth (Fig. 9CD). Finally, as might be expected the lack of *in vitro* CAE activity observed for ΔN_{20} TbCAE-myc and ΔN_{30} TbCAE-myc variants (Fig. 9C and D, right panels) correlates with their inability to complement the growth arrest (Fig 8, middle panels).

To determine the localization of the tagged TbCAE variants we used digitonin extraction as in Fig. 5A. The results in the left panel of Fig. 9A show that the full length protein is recovered in both the pellet and the supernatant fractions which is consistent with the protein being dually localized to the cytosol and possibly the

nucleus (both recovered in the supernatant fraction) as well as to the mitochondrion (recovered in the pellet). Interestingly, in contrast to the experiment shown Fig. 5A, we found a significant fraction of the protein in the pellet. However, it is important to consider that in the experiments shown in Fig. 9, in contrast to the one shown in Fig. 5A, the endogenous TbCAE is replaced with the TbCAE that is expressed from the recoded synthetic gene. Fig S1 shows that the latter is significantly overexpressed which could explain why it can now be detected in mitochondria. Complementation with the ΔN_{10} TbCAE-myc version of the protein (Fig. 9B) shows the same results than the full-length protein indicating that the N-terminal 10 amino acids are neither required for mitochondrial localization nor for enzymatic activity of TbCAE. However, the ΔN_{20} TbCAE-myc and ΔN_{30} TbCAE-myc variants, both did not restore growth and were enzymatically inactive (Fig. 9C and D). Interestingly, ΔN_{20} TbCAE-myc was exclusively cytosolically localized (Fig. 9C), whereas the ΔN_{30} TbCAE-myc variant was more efficiently imported than even the full-length protein (Fig. 9D). The lower panel in Fig. 9D shows that the pellet fraction of ΔN_{30} TbCAE-myc is fully soluble in 1% of digitonin indicating it is indeed imported and does not reflect large protein aggregates. In summary these results indicate that amino acids 11-20 are required for mitochondrial localization of TbCAE as well as for the CCA adding activity. Deleting a further 10 amino acids (21-30) then reveals a cryptic import signal that normally is buried within the protein (39) leading to an artefactual import of the truncated protein.

In summary, these results show that the N-terminal 10 amino acids are neither required for TbCAE activity nor for the mitochondrial localization of the protein. Further deletions however abolish both the mitochondrial localization as well as the enzymatic activity.

Complementation using yeast cytosolic CAE isoform

In order to determine whether the mitochondrial fraction of TbCAE is essential for growth, the TbCAE RNAi cell line was complemented by expressing the cytosolic/nuclear isoform of the CAE of *S. cerevisiae* (ScCAE). Fig. 10 shows that expression of the yeast enzyme restores growth

to nearly wildtype levels (Fig. 10AB). Northern analysis of total RNA isolated from the TbCAE RNAi cell lines reveals the expected (Fig. 3) accumulation of the slower migrating form of the tRNA^{lys} in the induced sample, which could be complemented by expression of ScCAE (Fig. 10C). In contrast, when mitochondrial RNAs were analyzed the accumulation of the slower migrating form of the tRNA^{lys} remained unaffected irrespectively of whether ScCAE was expressed or not (Fig. 10D). These results indicate that expression of the cytosolic/nuclear isoform of ScCAE in *T. brucei* selectively complements for the lack of the cytosolic fraction of TbCAE. However, since the complemented cell line is able to grow at near wild type levels this demonstrates that the mitochondrial localized TbCAE is dispensable for growth.

DISCUSSION

We have identified TbCAE as the single protein responsible for both nuclear/cytosolic as well as mitochondrial CCA adding activity. Experiments using an inducible TbCAE-RNAi cell line strongly suggest that TbCAE catalyzes the addition of CCA to 3'-cleaved tRNA precursors. Since only tRNAs carrying an intact CCA 3'-end can be aminoacylated TbCAE is expected to be essential. In line with this we observe that its ablation abolishes protein synthesis and cell growth. Moreover, it leads to accumulation of a subset of truncated tRNAs, although some tRNA isoacceptor groups are not affected, indicating that TbCAE may have different affinities for different tRNA isoacceptor groups. Interestingly, even though ablation of TbCAE inhibits the activity measured in total cellular as well as in mitochondrial extracts to essentially the same extent, imported mitochondrial tRNAs are much less affected than total tRNAs. In fact, accumulation of truncated mitochondrial forms at steady state could only be observed for the tRNA^{lys}. How can this be explained?

In trypanosomes, in contrast to most other systems, all mitochondrial tRNAs are imported from the cytosol. It has been demonstrated that binding to cytosolic EF1a is a prerequisite for a tRNA to be imported into mitochondria. However, EF1a itself is not imported into mitochondria, but mediates the essential targeting step (11). Since elongation factors

only recognize aminoacylated tRNAs (40) this implies that only charged tRNAs can be imported. This was independently confirmed in a recent publication showing that, while a mutant *E. coli* tRNA^{tyr} expressed in *T. brucei* could not be imported into mitochondria, because it was not recognized by the trypanosomal tyrosyl-tRNA synthetase, its import could be induced by co-expression of the *E. coli* tyrosyl-tRNA synthetase (12). These two examples illustrate that in *T. brucei* only aminoacylated tRNAs are imported into mitochondria in vivo. Since only tRNAs with a mature CCA ends can be aminoacylated, it follows that only fully processed tRNAs carrying an intact CCA are imported into mitochondria.

This indicates that the mitochondrial TbCAE may have a different function than its cytosolic/nuclear counterpart. It does not need to add the CCA-tail to immature cleaved tRNAs since they are not present in mitochondria. Instead, it might be involved in repair of damaged 3' ends. Such damage can occur by spontaneous or enzymatic hydrolysis, e. g. by one of the numerous ribonucleases that were detected in the *T. brucei* mitochondrion. The accumulation of truncated newly synthesized variant of tRNA^{met} (tRNA^{met}) after TbCAE knock down demonstrates that the CCA-ends of imported tRNAs are indeed turned over.

Expression of the yeast CAE isoform in the induced TbCAE RNAi cell line allows to selectively complement the extra-mitochondrial TbCAE activity. In the resulting cell line, that is devoid of mitochondrial CAE activity, growth is restored to nearly wildtype levels illustrating that *T. brucei* can grow in the absence of a TbCAE-based mitochondrial tRNA repair mechanism. However, the fact that the complemented cell line grows a little bit slower suggest that even when kept in culture mitochondrial TbCAE provides a slight fitness advantage to the cells. Trypanosomes have a complex life cycle and mitochondrial translation is known to be essential in both the procyclic and the bloodstream forms (4). Mitochondrial TbCAE activity might therefore well be essential under certain conditions explaining why trypanosomes have maintained mitochondrial CAE activity, even though the organelle imports mature tRNAs from the cytosol.

A similar situation has been described for *E. coli* in which deletion of the CAE resulted in a slow growth phenotype (41,42). Since all *E. coli* tRNA genes encode the CCA-tail this is likely caused by a deficient tRNA repair mechanism.

All eukaryotic CAEs investigated so far are targeted to multiple compartments. The yeast enzyme is the best studied one. Its mRNA has three in frame AUG codons upstream of the open reading frame for the CAE. Moreover, transcription creates mRNAs with heterogeneous 5'-ends. The enzyme variant synthesized from the most upstream AUG contains an N-terminal mitochondrial targeting signal and is imported into mitochondria, whereas the two shorter variants provide for the nuclear/cytosolic activities of the enzyme (32,43). In plants the problem is even more complex and the single CAE is targeted to the nucleus/cytosol as well as to mitochondria and plastids. Similar to yeast multiple in frame upstream start codons influence the localization of the enzyme. However, the stability of the mature part of the enzyme is also important for the intracellular distribution of the enzyme (34,35). We did not find any in frame AUG codons upstream of the TbCAE ORF suggesting that the intracellular localization of the enzyme is not regulated by different translation start sites. While differential transsplicing was shown to be responsible for dual targeting of trypanosomal isoleucyl-tRNA synthetase (44) there is no evidence for upstream alternative splice sites in the case of TbCAE. This makes differential trans-splicing an unlikely mechanism for differential localization. Our complementation experiments showed that amino acids 10-20 are required for mitochondrial targeting as well as for enzymatic activity. However, unlike a typical presequence, this segment cannot be cleaved off since this would inactivate the enzyme. Thus, it remains unclear how the multiple localization of TbCAE is regulated. One possibility is that, as has been shown for fumarase in yeast, the protein folding vs. translocation rate determines how much of the protein is imported into mitochondria (45).

Finally, it has been shown that the single imported tRNA^{Leu} in yeast mitochondria is co-imported with the precursor of the mitochondrial lysyl-tRNA synthetase (46). Since in trypanosomes a fraction of all but two

tRNAs are imported into mitochondria the partially imported TbCAE, which recognizes all tRNAs, appears to be an excellent candidate to piggyback tRNAs into the mitochondrion. However, the fact that ablation of TbCAE did not diminish the levels of any imported tRNAs and the same was the case for the fully viable cell line complemented with the cytosolic/nuclear isoform ScCAE demonstrates that TbCAE is not involved in tRNA import. The same has been shown previously for mitochondrial elongation factor Tu and various aaRSs. These results are therefore in line with a recent study showing that, in contrast to yeast, mitochondrial tRNAs in trypanosomes, while relying on some subunits of the protein import machinery, are not co-imported with proteins (9).

MATERIAL AND METHODS

Transgenic cell lines

Transgenic *T. brucei* cell lines are based on the procyclic strain 29-13 (47) and grown at 27°C in SDM-79 containing 10% (vol/vol) fetal calf serum (FCS) (Sigma). All the growth curves were performed in triplicates. Tetracycline (1 µg/ml; Sigma, Cat. No. T7660) was added on day 0 in the indicated +Tet cells.

The inducible TbCAE RNAi cell line was produced using a pLew100-derived plasmid whose resistance cassette was changed from phleomycin to blasticidine. The RNAi targets the nucleotides 300-856 of the ORF of TbCAE (Tb927.9.8780). The stem loop DNA fragment was produced by cloning the 300-856 TbCAE gene fragment in opposing directions separated by a 460 bp stuffer DNA.

To produce the construct expressing the C-terminal triple c-myc tagged version of the TbCAE, the respective ORF with the desired flanking restriction sites was amplified by PCR. The resulting PCR product was then ligated into a modified pLew100-based vector carrying a puromycin resistance and allowing the addition of the triple c-myc tag sequence at the C-terminus.

For complementation experiments, a synthetic gene of TbCAE was used (Biomatik). The codons between positions 300-856 of the TbCAE gene were changed such that its transcript would be RNAi resistant but still translate into the same amino acid sequence than found in the endogenous protein. The

sequence of the synthetic TbCAE gene is shown in the Fig. S4. Subsequently the synthetic gene was cloned into a pLew100-based vector allowing the addition of the triple c-myc tag at the C-terminus and transfected into the TbCAE RNAi cell line. To produce the constructs allowing expression of the N-terminally truncated TbCAE variants that were used for complementation experiment, the corresponding DNA fragments were amplified from the synthetic gene.

For complementation with the yeast CCA adding enzyme, the ORF of the *Saccharomyces cerevisiae cca1* gene from the yeast strain BY4742 was amplified starting from the 2nd in-frame ATG codon (at 28 bp) since this translates into the cytosolic/nuclear localized version of the protein (43). The resulting PCR product was cloned into the pLew100 based vector for C-terminal triple c-myc tagging.

Transfection of plasmids was done as described earlier (48). Expression was induced by adding tetracycline to a concentration of 1 μ g/ml to the culture medium.

Antibodies

Commercially available monoclonal antibodies used and their working dilutions for immunofluorescence (IF) and immunoblotting (IB) are as follows: mouse c-Myc (Invitrogen, No. 132500; IB 1:2000, IF 1:50), and mouse EF1a (Merck Millipore, No. 05-235; IB 1:10,000). Anti-VDAC, anti-ATOM40 and anti-CoxIV polyclonal antibodies were produced in rabbits and described previously (49,50). Working dilutions for immunoblots (IB) and immunofluorescence (IF) are: VDAC (IB 1:1000) and ATOM40 (IB 1:10,000; IF 1:1000). Mouse anti-TbVCP antibody was a generous gift from James D Bangs and has been described previously (51).

Commercially available secondary antibodies for IB are IRDye 680LT goat anti-mouse, IRDye 800CW goat anti-rabbit (LI-COR Biosciences, 1:20,000). Secondary antibodies used for IF are goat anti-mouse Alexa Fluor 596 and goat anti-rabbit Alexa Fluor 488 (ThermoFisher Scientific, 1:1000).

Labelling of proteins with ³⁵S-methionine

5x10⁶ cells from an exponentially growing culture were washed with PBS and resuspended in 400 μ l of SDM-80 medium (52) lacking L-methionine. Subsequently, the suspension was

supplemented with 2 μ l of ³⁵S-methionine (10 μ Ci/ μ l, specific activity 1000Ci/mmol, Hartmann analytics). The cells were incubated at 27°C for 20 minutes after which they were washed with PBS and resuspended in sodium dodecylsulfate (SDS)-PAGE sample buffer. In the negative control, cycloheximide (300 μ g/ml final concentration) was added to the culture one minute before the addition of ³⁵S-methionine. Proteins were denatured at 95°C for 5 minutes, subjected to SDS-PAGE and subsequently stained with Coomassie. The gel was dried, and radioactive signal was analysed using a Phosphorimager.

CCA adding enzyme assay

2x10⁷ cells from an exponentially growing cultures of uninduced and induced TbCAE RNAi cell line were washed with PBS and resuspended in 50 μ l in CCA-buffer (30mM HEPES/KOH pH 7.6, 30mM KCl, 6mM MgCl₂) containing 0.1% Triton X-100 and incubated at 4°C for 15 minutes. Subsequently, the samples were centrifuged at 16,000xg in a table top centrifuge and the supernatants corresponding to the total cellular extract were collected. Next 5 μ L each of the extracts and 2 μ g of purified total RNA isolated from wild-type *T. brucei* cells were mixed with CCA buffer containing 10 μ M of DTT, subsequently unlabelled ATP (or CTP) (0.1mM final) and α -³²P-CTP (or α -³²P-ATP or α -³²P-UTP) (10 μ Ci, specific activity 3000Ci/mmol; Hartmann analytics) were added and the reaction was adjusted to 20 μ l.

The reactions were incubated at 27°C for 1 hour and RNA was extracted using the acid guanidinium method (53). The extracted RNA samples were subjected to 8M Urea/15% PAGE, the gel was stained with ethidium bromide, dried and analysed using a Phosphorimager.

To determine the mitochondrial TbCAE activity mitochondria were isolated from a culture of the uninduced and the 2.5 days induced TbCAE RNAi cell line as described (54). Same protein equivalents of isolated mitochondria of each culture were washed with PBS and lysed in CCA buffer containing 0.5% of Triton X-100 and incubated for 15 minutes at 4°C. To 50 μ l of extracts unlabelled ATP (0.1mM final) and α -³²P-CTP (10 μ Ci, specific activity 3000Ci/mmol; Hartmann analytics) were added, incubated for 1 hour at 27°C before

the RNA was extracted using acid guanidium method (53). The extracted RNA was subjected to 8M Urea/15% PAGE, stained with ethidium bromide and processed as described above.

RNA extraction and northern analysis

For analysis of total tRNAs, uninduced and induced TbCAE RNAi cells were harvested and washed with PBS before the total RNA was extracted using acid guanidium method (53). For isolation of the corresponding mitochondrial RNAs, the cells were permeabilized using low digitonin concentration (0.025%) and the contaminating cytosolic RNA was degraded using RNase A as described (38). Subsequently RNA was isolated from the resulting crude mitochondrial fractions by the acid guanidium method (53). 5 μ g of total RNA or 2.5 μ g of mitochondrial RNA each were separated on an 8M Urea/15% PAGE and visualised using ethidium bromide staining. Subsequently gels were electroblotted onto a charged nylon membrane using semi-dry blotting apparatus (300mAmps, 1h, Biorad) and probed with the corresponding labelled oligonucleotide (Table S1). The blots were hybridized in 6x SSPE (60 mM NaH₂PO₄, 0.9 M NaCl, 6 mM EDTA) containing 5x Denhardt's and 0.5% (w/v) SDS at 55 °C for 14–18 h using 10 pmols of the corresponding 5'-labelled oligonucleotide. Labelling of the oligonucleotides was done with γ -³²P-ATP (50 μ Ci specific activity 3000Ci/ mmol; Hartmann analytics) using the polynucleotide kinase forward reaction. The Northern blots were analysed using a Phosphorimager.

For mRNA Northern blots, 7 μ g of total RNA was separated on a 1% agarose gel in 20mM MOPS buffer, pH 7.0, containing 0.5% formaldehyde. The probe used for the Northern analysis corresponds to the fragment

(nucleotides 300-856) of the TbCAE ORF which was gel-purified from a PCR product and radioactively labelled using the Prime-a-Gene labelling kit (Promega).

Digitonin extraction

1x10⁶ of exponentially growing cells were incubated for 10 minutes on ice in 0.025% (w/v) digitonin made up in 1X SoTE (20mM Tris-HCl pH 7.5, 0.6 M sorbitol, 2mM EDTA). These conditions selectively lyse the cell membrane but leave the mitochondrial membranes intact. After centrifugation in a table top centrifuge at 6,800xg, 4°C, the resulting crude mitochondrial pellet was separated from the supernatant, corresponding to a crude cytosolic fraction. Total cells, crude cytosolic and mitochondrial fractions were mixed with SDS sample buffer and 2.5x10⁶ cell equivalents of each fraction were subjected to 12% SDS-PAGE and immunoblotting.

Immunofluorescence

TbCAE-myc expression was induced for 24 hours using tetracycline (1 μ g/ml). Cells were fixed with 4% paraformaldehyde in 1X PBS, washed with 1X PBS and allowed to adhere on a slide. Subsequently the cells were permeabilized on the slide with PBS containing 1% of Triton-X100 for 2 minutes after which primary antibody diluted in PBS was added for 1 hour. The slides were washed three times with PBS and incubated with the corresponding secondary antibodies. After three more washes with PBS, the slides were mounted using VectaShield containing 4,6-diamidino- 2-phenylindole (DAPI) (Vector Laboratories, P/N H-1200). Images were taken with a DFC360 FX monochrome camera (Leica Microsystems) mounted on a DMI6000B microscope (Leica Microsystems). Image analysis was done using LAS X software (Leica Microsystems).

DATA AVAILABILITY STATEMENT: All data are contained within the manuscript.

ACKNOWLEDGMENTS

We would like to thank Dr Moritz Niemann for his help with the mitoprep. This study was supported by the NCCR "RNA & Disease" and in part by grants 175563 both funded by the Swiss National Science Foundation.

CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest with the contents of this article.

REFERENCES

1. Shikha, S., Brogli, R., Schneider, A., and Polacek, N. (2019) tRNA Biology in Trypanosomes. *Chimia* **73**, 395-405
2. Alfonzo, J. D., and Söll, D. (2009) Mitochondrial tRNA import--the challenge to understand has just begun. *Biol. Chem.*, 717-722
3. Niemann, M., Schneider, A., and Cristodero, M. (2011) Mitochondrial translation in trypanosomatids: a novel target for chemotherapy? *Trends Parasitol.* **27**, 429-433
4. Cristodero, M., Seebeck, T., and Schneider, A. (2010) Mitochondrial translation is essential in bloodstream forms of *Trypanosoma brucei*. *Mol. Microbiol.* **78**, 757-769
5. Schneider, A. (2011) Mitochondrial tRNA Import and Its Consequences for Mitochondrial Translation. *Ann. Rev. Biochem.* **80**, 1033-1053
6. Sieber, F., Duchene, A. M., and Marechal-Drouard, L. (2011) Mitochondrial RNA import: from diversity of natural mechanisms to potential applications. *International review of cell and molecular biology* **287**, 145-190
7. Salinas, T., Duchene, A. M., and Marechal-Drouard, L. (2008) Recent advances in tRNA mitochondrial import. *Trends Biochem Sci* **33**, 320-329
8. Tschopp, F., Charriere, F., and Schneider, A. (2011) In vivo study in *Trypanosoma brucei* links mitochondrial transfer RNA import to mitochondrial protein import. *EMBO Rep* **12**, 825-832
9. Niemann, M., Harsman, A., Mani, J., Peikert, C. D., Oeljeklaus, S., Warscheid, B., Wagner, R., and Schneider, A. (2017) tRNAs and proteins use the same import channel for translocation across the mitochondrial outer membrane of trypanosomes. *Proc Natl Acad Sci U S A* **114**, E7679-E7687
10. Seidman, D., Johnson, D., Gerbasi, V., Golden, D., Orlando, R., and Hajduk, S. (2012) Mitochondrial membrane complex that contains proteins necessary for tRNA import in *Trypanosoma brucei*. *J. Biol. Chem.* **287**, 8892-8903
11. Bouzaidi-Tiali, N., Aeby, E., Charrière, F., Pusnik, M., and Schneider, A. (2007) Elongation factor 1a mediates the specificity of mitochondrial tRNA import in *T. brucei*. *EMBO J.* **26**, 4302-4312
12. Huot, J. L., Shikha, S., and Schneider, A. (2019) Inducible orthogonal aminoacylation demonstrates that charging is required for mitochondrial tRNA import in *Trypanosoma brucei*. *Sci Rep* **9**, 10836
13. Charrière, F., O'Donoghue, P., Helgadóttir, S., Maréchal-Drouard, L., Cristodero, M., Horn, E. K., Söll, D., and Schneider, A. (2009) Dual Targeting of a tRNA^{Asp} Requires Two Different Aspartyl-tRNA Synthetases in *Trypanosoma brucei*. *J. Biol. Chem.* **284**, 16210-16217
14. Charrière, F., Helgadóttir, S., Horn, E. K., Söll, D., and Schneider, A. (2006) Dual targeting of a single tRNA^{Trp} requires two different tryptophanyl-tRNA synthetases in *Trypanosoma brucei*. *Proc. Natl. Acad. Sci. USA* **103**, 6847-6852
15. Peikert, C. D., Mani, J., Morgenstern, M., Käser, S., Knapp, B., Wenger, C., Harsman, A., Oeljeklaus, S., Schneider, A., and Warscheid, B. (2017) Charting Organellar Importomes by Quantitative Mass Spectrometry. *Nat. Commun.* **8**, 15272
16. Alfonzo, J. D., Blanc, V., Estevez, A. M., Rubio, M. A. T., and Simpson, L. (1999) C to U editing of anticodon of imported mitochondrial tRNA^{Trp} allows decoding of UGA stop codon in *Leishmania*. *EMBO J.* **18**, 7056-7062
17. Paris, Z., and Alfonzo, J. D. (2018) How the intracellular partitioning of tRNA and tRNA modification enzymes affects mitochondrial function. *IUBMB Life*
18. Wohlgamuth-Benedum, J. M., Rubio, M. A., Paris, Z., Long, S., Poliak, P., Lukes, J., and Alfonzo, J. D. (2009) Thiolation controls cytoplasmic tRNA stability and acts as a negative determinant for tRNA editing in mitochondria. *J. Biol. Chem.* **284**, 23947-23953
19. Crain, P. F., Alfonzo, J. D., Rozenski, J., Kapushoc, S. T., McCloskey, J. A., and Simpson, L. (2002) Modification of the universally unmodified uridine-33 in a mitochondria-imported

- edited tRNA and the role of the anticodon arm structure on editing efficiency. *RNA* **8**, 752-761
20. Aeby, E., Palioura, S., Pusnik, M., Marazzi, J., Lieberman, A., Ullu, E., Söll, D., and Schneider, A. (2009) The canonical pathway for selenocysteine insertion is dispensable in Trypanosomes. *Proc. Natl. Acad. Sci. USA* **106**, 5088-5092
 21. Aeby, E., Seidel, V., and Schneider, A. (2009) The selenoproteome is dispensable in bloodstream forms of *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* **168**, 191-193
 22. Aeby, E., Ullu, E., Yepiskoposyan, H., Schimanski, B., Roditi, I., Mühlemann, O., and Schneider, A. (2010) tRNA^{Sec} is transcribed by RNA polymerase II in *Trypanosoma brucei* but not in humans. *Nucleic Acids Res.* **38**, 5833-5843
 23. Hartmann, R. K., Gossringer, M., Spath, B., Fischer, S., and Marchfelder, A. (2009) The making of tRNAs and more - RNase P and tRNase Z. *Prog Mol Biol Transl Sci* **85**, 319-368
 24. Maraia, R. J., and Lamichhane, T. N. (2011) 3' processing of eukaryotic precursor tRNAs. *Wiley interdisciplinary reviews. RNA* **2**, 362-375
 25. Rossmannith, W. (2012) Of P and Z: mitochondrial tRNA processing enzymes. *Biochim Biophys Acta* **1819**, 1017-1026
 26. Taschner, A., Weber, C., Buzet, A., Hartmann, R. K., Hartig, A., and Rossmannith, W. (2012) Nuclear RNase P of *Trypanosoma brucei*: a single protein in place of the multicomponent RNA-protein complex. *Cell Rep* **2**, 19-25
 27. Salavati, R., Panigrahi, A. K., and Stuart, K. D. (2001) Mitochondrial ribonuclease P activity of *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* **115**, 109-117
 28. Mörl, M., and Marchfelder, A. (2001) The final cut: The importance of tRNA 3'-processing. *EMBO J.* **2**, 17-20
 29. Betat, H., and Morl, M. (2015) The CCA-adding enzyme: A central scrutinizer in tRNA quality control. *Bioessays* **37**, 975-982
 30. Hancock, K., LeBlanc, A. J., Donze, D., and Hajduk, S. L. (1992) Identification of nuclear encoded precursor tRNAs within the mitochondrion of *Trypanosoma brucei*. *J. Biol. Chem.* **267**, 23963-23971
 31. Béja, O., Ullu, E., and Michaeli, S. (1993) Identification of a tRNA-like molecule that copurifies with the 7SL RNA of *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* **57**, 223-230
 32. Wolfe, C. L., Lou, Y. C., Hopper, A. K., and Martin, N. C. (1994) Interplay of heterogeneous transcriptional start sites and translational selection of AUGs dictate the production of mitochondrial and cytosolic/nuclear tRNA nucleotidyltransferase from the same gene in yeast. *J Biol Chem* **269**, 13361-13366
 33. Chen, J. Y., Joyce, P. B., Wolfe, C. L., Steffen, M. C., and Martin, N. C. (1992) Cytoplasmic and mitochondrial tRNA nucleotidyltransferase activities are derived from the same gene in the yeast *Saccharomyces cerevisiae*. *J Biol Chem* **267**, 14879-14883
 34. von Braun, S. S., Sabetti, A., Hanic-Joyce, P. J., Gu, J., Schleiff, E., and Joyce, P. B. (2007) Dual targeting of the tRNA nucleotidyltransferase in plants: not just the signal. *Journal of experimental botany* **58**, 4083-4093
 35. Leibovitch, M., Bublak, D., Hanic-Joyce, P. J., Tillmann, B., Flinner, N., Amsel, D., Scharf, K. D., Mirus, O., Joyce, P. B., and Schleiff, E. (2013) The folding capacity of the mature domain of the dual-targeted plant tRNA nucleotidyltransferase influences organelle selection. *Biochem J* **453**, 401-412
 36. Pusnik, M., Schmidt, O., Perry, A. J., Oeljeklaus, S., Niemann, M., Warscheid, B., Lithgow, T., Meisinger, C., and Schneider, A. (2011) Mitochondrial preprotein translocase of trypanosomatids has a bacterial origin. *Curr. Biol.* **21**, 1738-1743
 37. Harsman, A., Niemann, M., Pusnik, M., Schmidt, O., Burmann, B. M., Hiller, S., Meisinger, C., Schneider, A., and Wagner, R. (2012) Bacterial origin of a mitochondrial outer membrane protein translocase: new perspectives from comparative single channel electrophysiology. *J Biol Chem* **287**, 31437-31445
 38. Tan, T. H. P., Pach, R., Crausaz, A., Ivens, A., and Schneider, A. (2002) tRNAs in *Trypanosoma brucei*: genomic organization, expression and mitochondrial import. *Mol. Cell. Biol.* **22**, 3707-3717

39. Hurt, E. C., and Schatz, G. (1987) A cytosolic protein contains a cryptic mitochondrial targeting signal. *Nature* **325**, 499-503
40. Ribeiro, S., Nock, S., and Sprinzl, M. (1995) Purification of aminoacyl-tRNA by affinity chromatography on immobilized *Thermus thermophilus* EF-Tu.GTP. *Anal. Biochem.* **228**, 330-335
41. Zhu, L., and Deutscher, M. P. (1987) tRNA nucleotidyltransferase is not essential for *Escherichia coli* viability. *EMBO J* **6**, 2473-2477
42. Reuven, N. B., Zhou, Z., and Deutscher, M. P. (1997) Functional overlap of tRNA nucleotidyltransferase, poly(A) polymerase I, and polynucleotide phosphorylase. *J Biol Chem* **272**, 33255-33259
43. Wolfe, C. L., Hopper, A. K., and Martin, N. C. (1996) Mechanisms leading to and the consequences of altering the normal distribution of ATP(CTP):tRNA nucleotidyltransferase in yeast. *J. Biol. Chem.* **271**, 4679-4686
44. Rettig, J., Wang, Y., Schneider, A., and Ochsenreiter, T. (2012) Dual targeting of isoleucyl-tRNA synthetase in *Trypanosoma brucei* is mediated through alternative trans-splicing. *Nucleic Acids Res.* **40**, 1299-1306
45. Regev-Rudzki, N., Yogev, O., and Pines, O. (2008) The mitochondrial targeting sequence tilts the balance between mitochondrial and cytosolic dual localization. *J Cell Sci* **121**, 2423-2431
46. Tarassov, I., Entelis, N., and Martin, R. P. (1995) Mitochondrial import of a cytoplasmic lysine-tRNA in yeast is mediated by cooperation of cytoplasmic and mitochondrial lysyl-tRNA synthetases. *EMBO J.* **14**, 3461-3471
47. Wirtz, E., Leal, S., Ochatt, C., and Cross, G. A. (1999) A tightly regulated inducible expression system for conditional gene knock-outs and dominant-negative genetics in *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* **99**, 89-101
48. McCulloch, R., Vassella, E., Burton, P., Boshart, M., and Barry, J. D. (2004) Transformation of monomorphic and pleomorphic *Trypanosoma brucei*. *Methods Mol. Biol.* **262**, 53-86
49. Niemann, M., Wiese, S., Mani, J., Chanfon, A., Jackson, C., Meisinger, C., Warscheid, B., and Schneider, A. (2013) Mitochondrial outer membrane proteome of *Trypanosoma brucei* reveals novel factors required to maintain mitochondrial morphology. *Mol. Cell. Proteomics* **12**, 515-528
50. Mani, J., Desy, S., Niemann, M., Chanfon, A., Oeljeklaus, S., Pusnik, M., Schmidt, O., Gerbeth, C., Meisinger, C., Warscheid, B., and Schneider, A. (2015) Mitochondrial protein import receptors in Kinetoplastids reveal convergent evolution over large phylogenetic distances. *Nat Commun* **6**, 6646
51. Roggy, J. L., and Bangs, J. D. (1999) Molecular cloning and biochemical characterization of a VCP homolog in African trypanosomes. *Mol Biochem Parasitol* **98**, 1-15
52. Lamour, N., Riviere, L., Coustou, V., Coombs, G. H., Barrett, M. P., and Bringaud, F. (2005) Proline metabolism in procyclic *Trypanosoma brucei* is down-regulated in the presence of glucose. *J. Biol. Chem.* **280**, 11902-11910
53. Chomczynski, P., and Sacchi, N. (1987) Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* **162**, 156-159
54. Schneider, A., Charrière, F., Pusnik, M., and Horn, E. K. (2007) Isolation of mitochondria from procyclic *Trypanosoma brucei*. *Methods Mol. Biol.* **372**, 67-80

The abbreviations used are: aaRSs, Aminoacyl-tRNA synthetases; ATOM40, Atypical translocase of the outer membrane 40; BLAST, Basic local alignment search tool; CAE, CCA-adding enzyme; DAPI, 4,6-diamidino- 2-phenylindole; EF1a, Eukaryotic elongation factor 1a; FCS, Fetal calf serum; SDM-79, Semi-defined medium-79; TbCAE: *Trypanosoma brucei* CCA-adding enzyme; TbVCP, *Trypanosoma brucei* valosin containing protein; VDAC, Voltage-dependent anion channel.

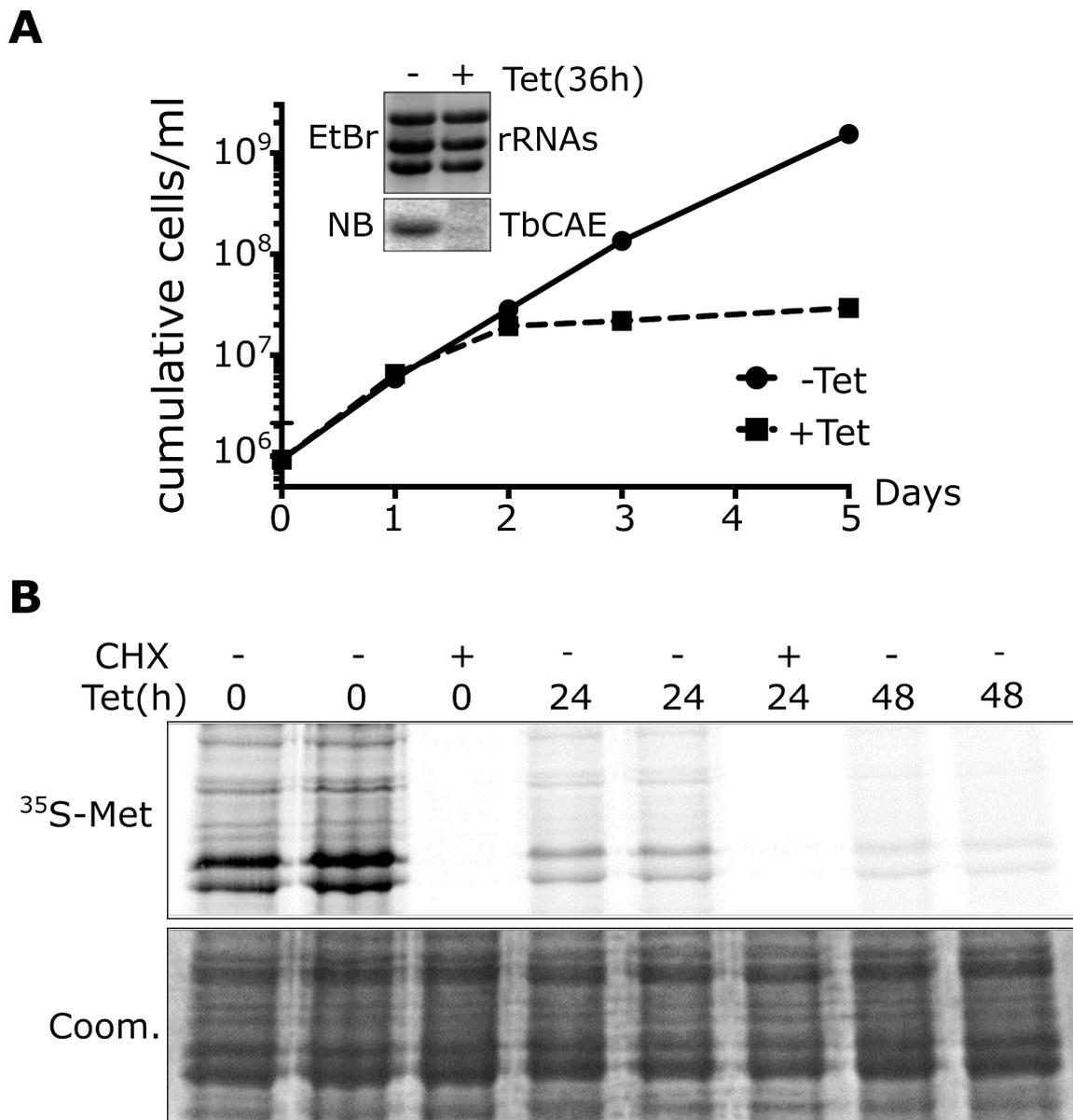


Figure 1 | TbCAE is essential for cytosolic translation. (A) Growth curve of uninduced (-tet) and induced (+tet) TbCAE RNAi cell line. The experiment has been performed in triplicates; the very small standard deviations are indicated. Insets show Northern blots (NB) of total RNA from uninduced or induced cells which were probed for TbCAE mRNA. Ethidium bromide stained rRNAs serve as loading controls. (B) ³⁵S-methionine labelling of the TbCAE RNAi cell line after 0, 24 and 48 hours of RNAi induction. Duplicate samples are shown. The corresponding coomassie stained gel serves as loading control. Cycloheximide (CHX) (300 μg/ml) is added to the negative control sample.

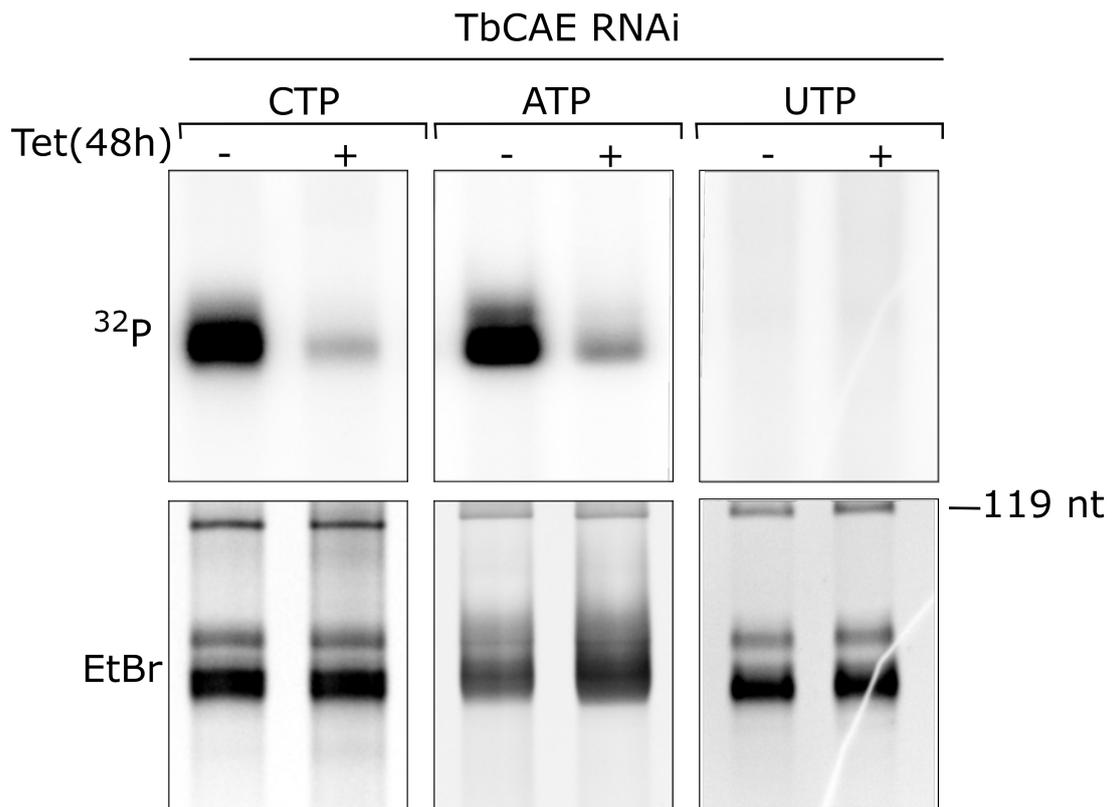


Figure 2 | Ablation of TbCAE inhibits its activity. Incorporation of ³²P-CTP, ³²P-ATP and ³²P-UTP into tRNAs was assayed in whole cell extracts prepared from the uninduced (-tet) and induced (+tet) TbCAE RNAi cell line. Only the tRNA regions of the gel are shown. The ethidium bromide (EtBr) stained gel serves as a loading control.

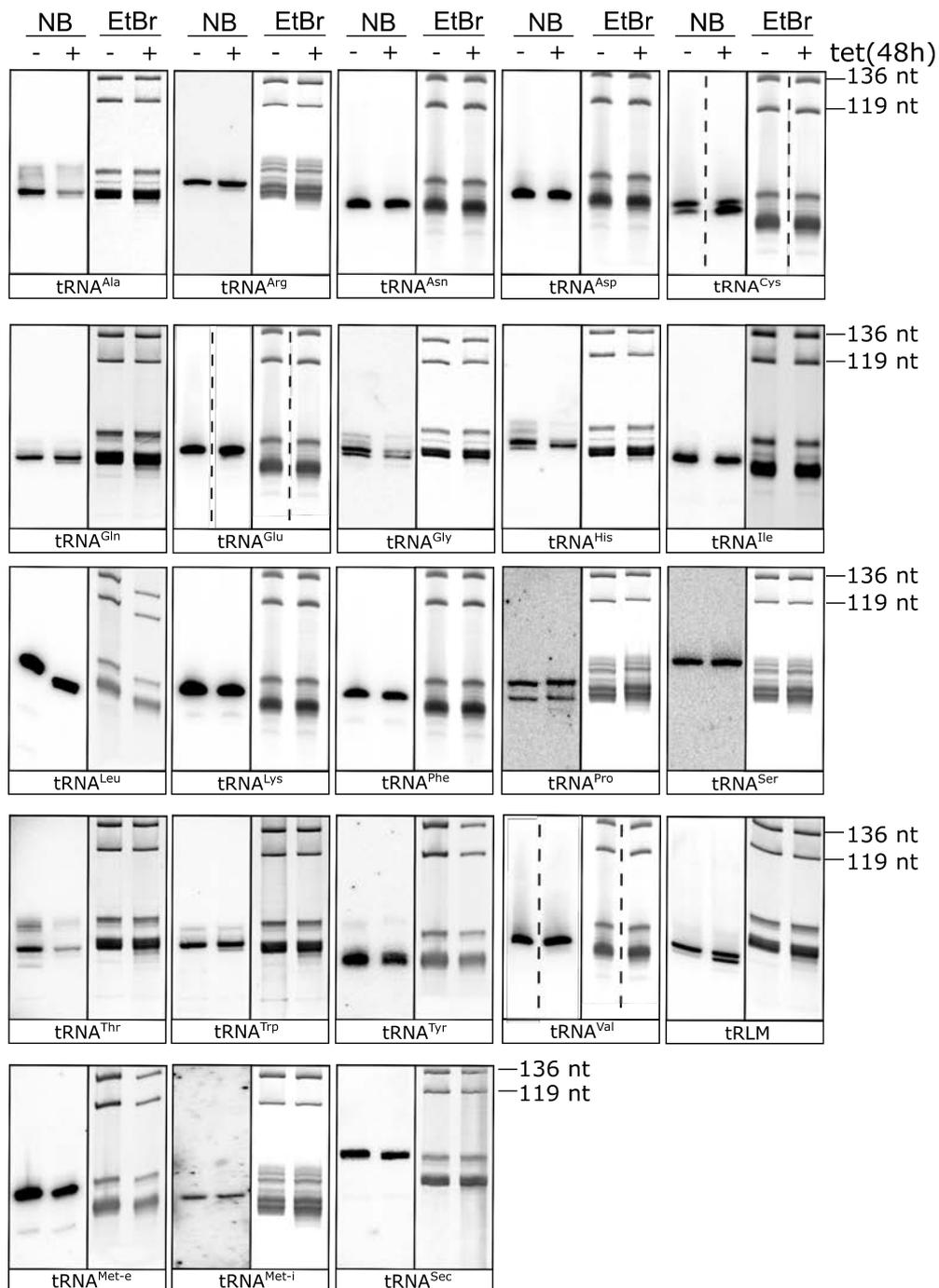


Figure 3 | Ablation of TbCAE differentially affects different tRNAs. Northern blots (NB) of total cellular RNA isolated from the uninduced (-) and induced (+) TbCAE RNAi cell line probed by specific oligonucleotide hybridization, for all tRNA isoacceptor groups and the tRNA-like molecule (tRLM). Only the tRNA regions of the blots and ethidium bromide (EtBr) stained gels are shown. One blot has been reprobated. Thus, the loading controls (EtBr-stained gels) for tRNA^{Cys} and tRNA^{Gln} are identical. For oligonucleotides used for hybridization, see Table S1.

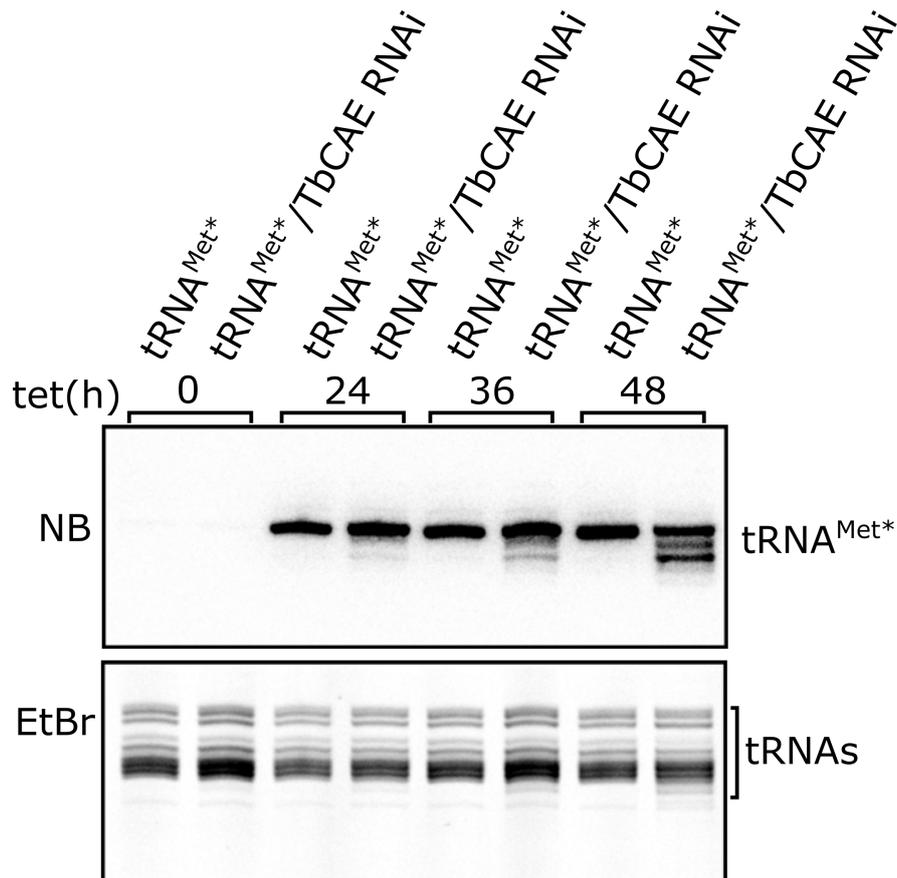


Figure 4 | TbCAE RNAi causes accumulation of newly synthesized truncated $tRNA^{Met^*}$. Total RNA isolated from cell lines allowing either tetracycline-inducible expression of variant $tRNA^{Met^*}$ ($tRNA^{Met^*}$) only, or inducible expression of $tRNA^{Met^*}$ in combination with RNAi against TbCAE were analysed by Northern blots at the indicated time points. The newly synthesized $tRNA^{Met^*}$ was detected by specific oligonucleotide hybridization. Only the tRNA region of the gel is shown. The ethidium bromide (EtBr) stained gel serves as a loading control.

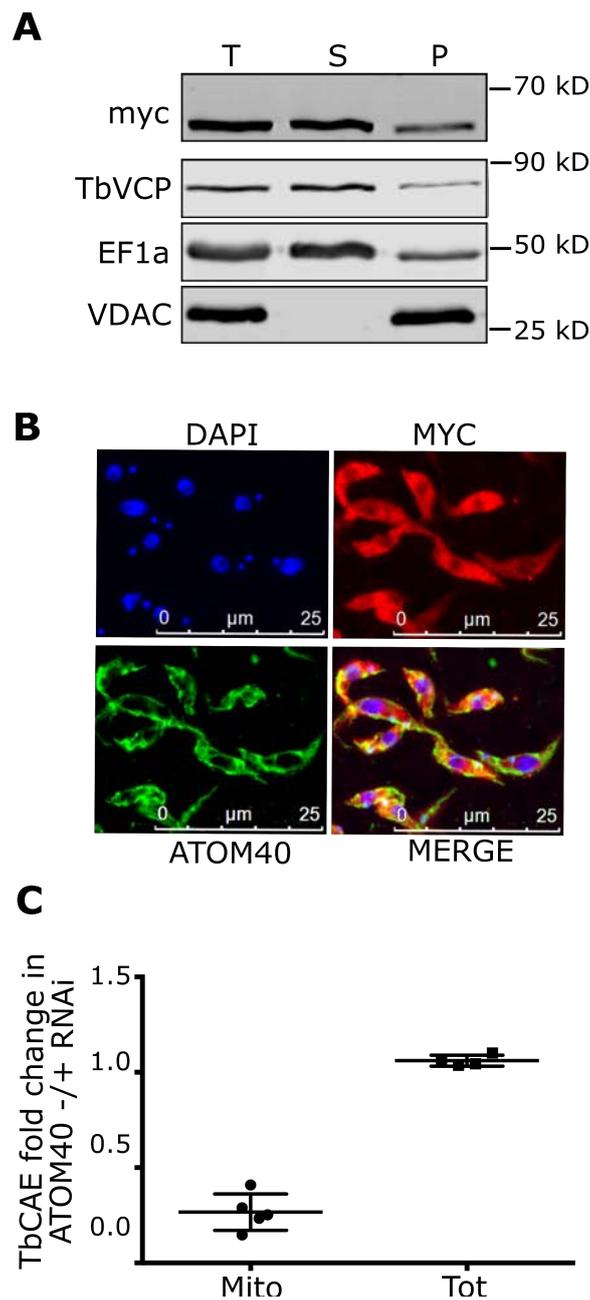


Figure 5 | TbCAE is dually localised in cytosol/nucleus and inside mitochondria. (A) Digitonin extracted total cells expressing triple c-myc tagged TbCAE (T) to separate a mitochondria-enriched fraction (P) from the cytosol-containing supernatant (S). EF1a and TbVCP are used as a cytosolic and VDAC as a mitochondrial marker, respectively. (B) Immunofluorescence analysis of whole cells expressing triple c-myc tagged TbCAE probed with anti-myc (red) and ATOM40 (green) antibodies. ATOM40 serves as a mitochondrial marker, nuclear and kinetoplast DNA is stained with DAPI (blue). (C) Fold change of TbCAE amounts in isolated mitochondria (Mito) and total cells (Tot) from uninduced and induced ATOM40 RNAi cell lines, respectively depicted as scatter dot plot. Data are from a previously published SILAC-based proteomic analysis (15).

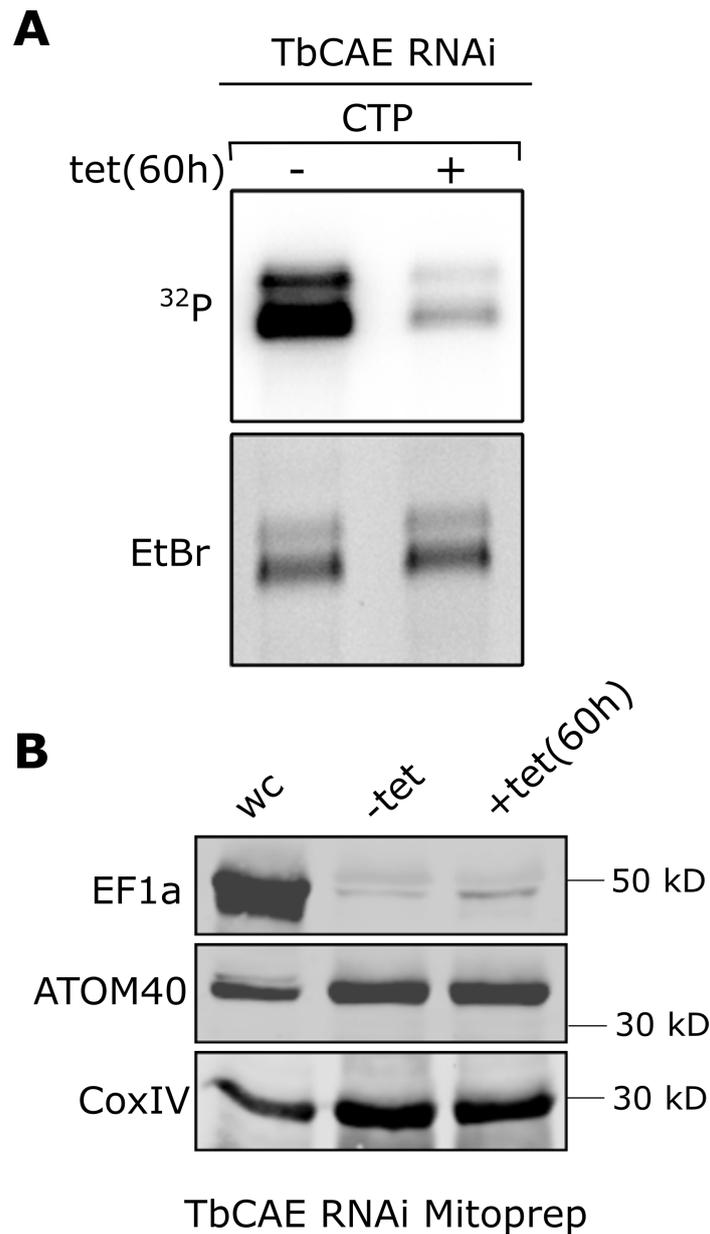


Figure 6 | TbCAE RNAi inhibits mitochondrial CAE activity. (A) Incorporation of ³²P-CTP into tRNAs was assayed in mitochondrial extracts prepared from the uninduced (-tet) and induced (+tet) TbCAE RNAi cell line. Only the tRNA region of the gels are shown. The ethidium bromide (EtBr) stained gel served as a loading control. (B) The total cellular extract (WC) and the mitochondrial (Mito) extracts used in (A) were analysed by immunoblots using a cytosolic (EF1a) and mitochondrial markers (ATOM40, CoxIV), respectively.

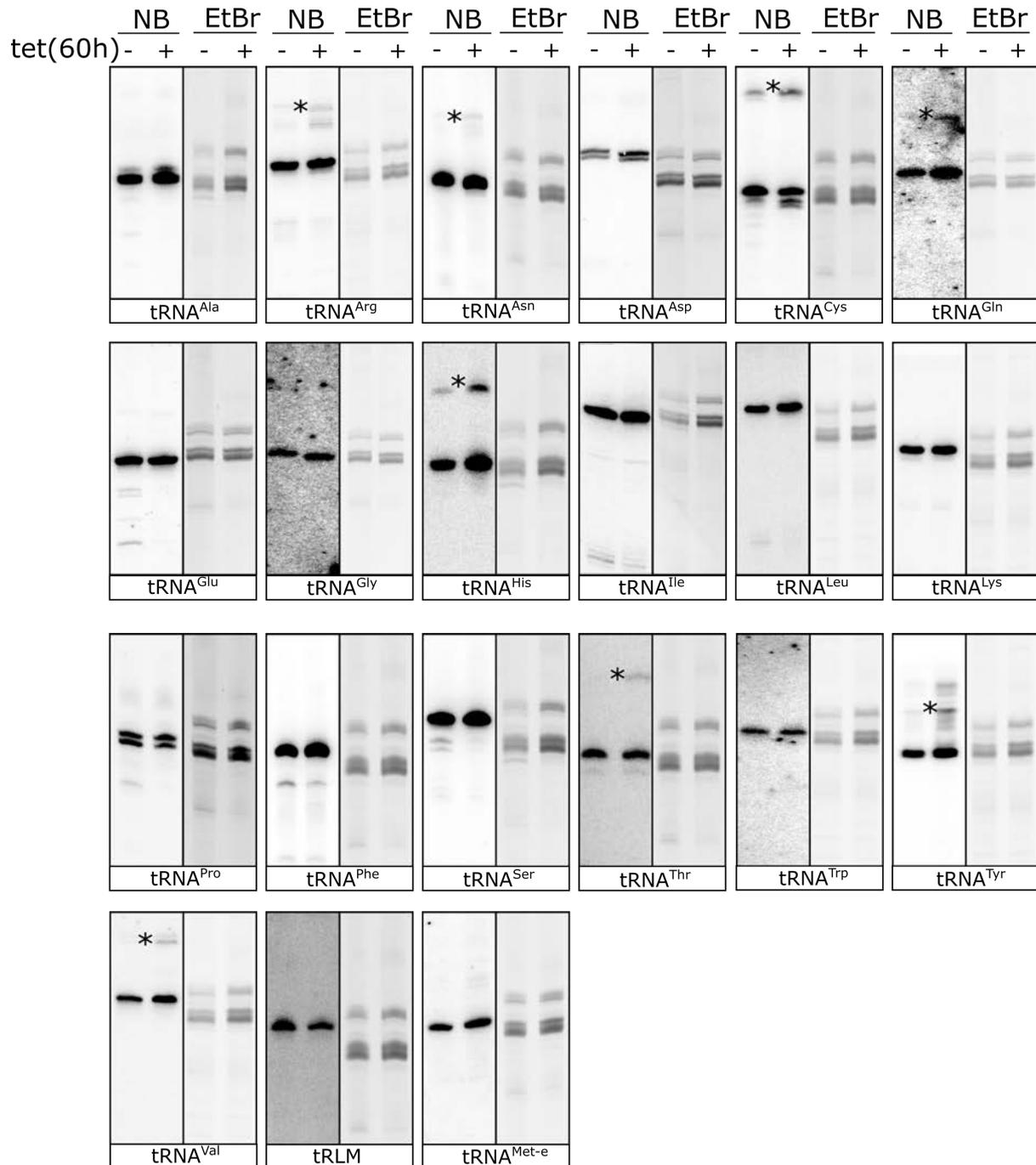


Figure 7 | TbCAE RNAi only moderately affects mitochondrial tRNAs. Northern blots (NB) of mitochondrial RNA isolated from the uninduced (-) and induced (+) TbCAE RNAi cell line probed by specific oligonucleotide hybridization for all tRNA isoacceptor groups and the tRNA-like molecule (tRLM). Only the tRNA regions of the blots and the gels are shown. The ethidium bromide (EtBr) stained gel serves as loading controls. Some blots have been reprobed. In these cases the corresponding loading controls (EtBr-stained gels) are identical. This applies for: tRNA^{Asp} and tRNA^{Gly}; tRNA^{Glu} and tRNA^{Gln}; tRNA^{Cys} and tRNA^{Thr}; tRNA^{Leu}, tRNA^{Lys} and tRNA^{Tyr}; tRNA^{Val} and tRNA^{Trp}. For oligonucleotides used for hybridization see Table S1. *Circular derivatives of tRNAs caused by the action of tRNA ligase present in the mitochondria.

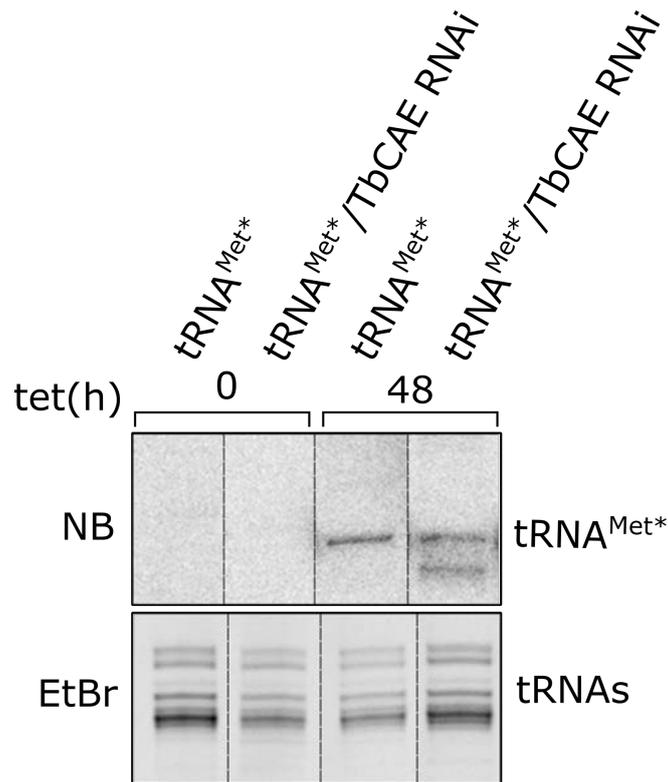


Figure 8 | TbCAE RNAi causes accumulation of newly synthesized truncated tRNA^{Met*} variant in mitochondria. Mitochondrial RNAs isolated from cell lines allowing either tetracycline-inducible expression of variant tRNA^{Met*} (tRNA^{Met*}) only, or inducible expression of the tRNA^{Met*} in combination with TbCAE RNAi were analysed by Northern blots (NB) at the indicated time points. The newly synthesized tRNA^{Met*} was detected by specific nucleotide hybridization. Only the tRNA region of the gel is shown. The ethidium bromide (EtBr) stained gel serves as a loading control.

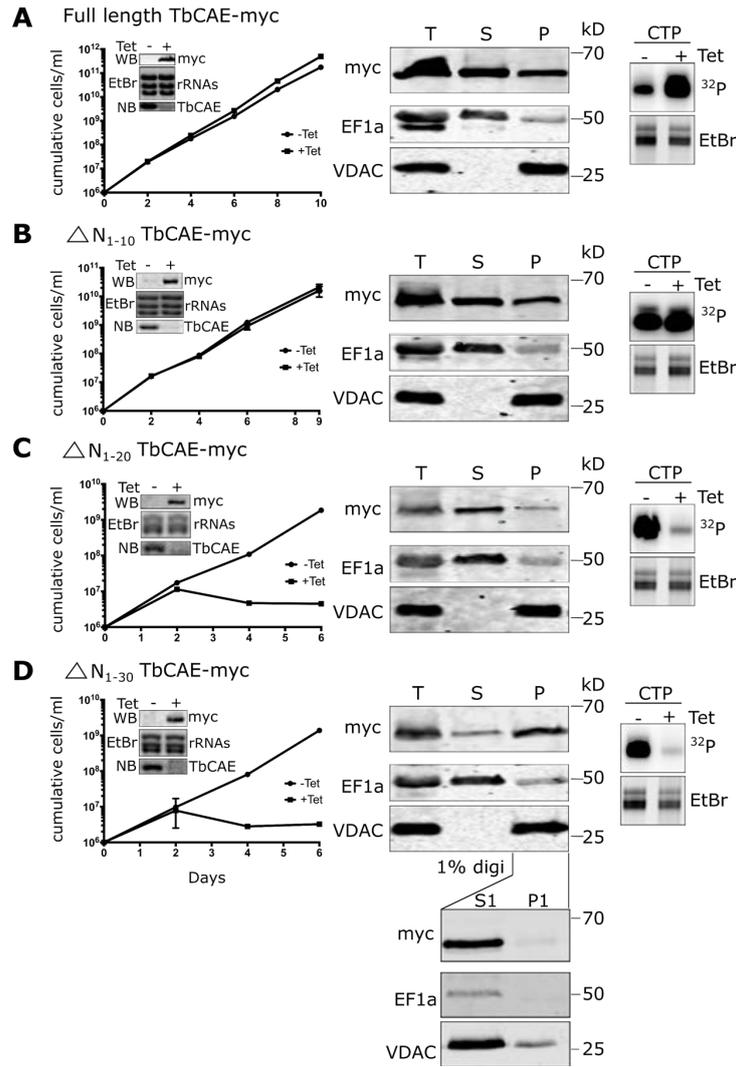


Figure 9 | Analysis of function and localization of TbCAE variants by complementation experiments. The TbCAE RNAi cell line was complemented by ectopic expression of C-terminally myc-tagged full length TbCAE derived from a synthetic gene (A), or derivatives thereof lacking the N-terminal 10 (B), 20 (C) or 30 amino acids (D). The constructs were termed ΔN_{1-10} TbCAE-myc, ΔN_{1-20} TbCAE-myc and ΔN_{1-30} TbCAE-myc respectively. Left panels, growth curves of the indicated uninduced (-tet) and induced (+tet) complemented cell lines. Insets in left panel: immunoblots of the myc-tagged TbCAE variant; loading control showing ethidium bromide-stained (EtBr) tRNAs; Northern blots of total RNA from uninduced (-) and induced (+) cells probed for TbCAE mRNA after 48 hours of tetracycline induction. Middle panels, digitonin extractions (0.025%) of the indicated induced complemented cell line. Ectopically expressed TbCAE was detected in total (T), mitochondria-enriched (P) and cytosol-containing supernatant (S) using anti-myc antibody. EF1a is used as a cytosolic and VDAC as a mitochondrial marker, respectively. Right panels, incorporation of 32 P-CTP into tRNAs was assayed in total cell extracts prepared from the indicated uninduced (-tet) and induced (+tet) complemented cell lines. Only the tRNA region of the gels are shown. The ethidium bromide (EtBr) stained gels serves as a loading control. The bottom panel in (D) shows that ΔN_{1-30} TbCAE-myc that co-fractionates with the mitochondria-enriched pellet (P) is fully soluble in 1% digitonin and is retained completely in the soluble mitochondrial fraction (S1) instead of the insoluble mitochondrial fraction (P1).

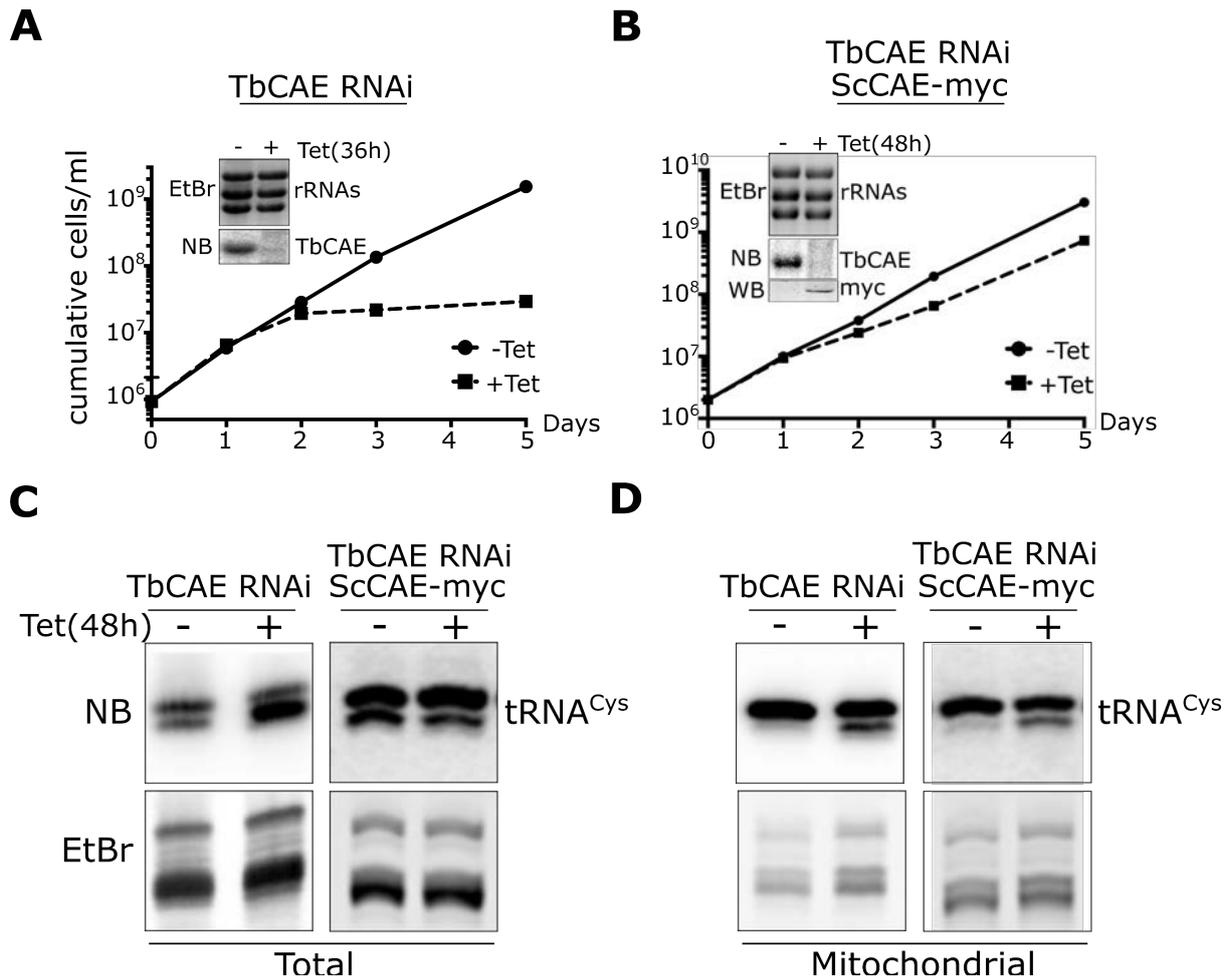


Figure 10 | Complementation of the TbCAE RNAi cell line by yeast cytosolic/nuclear ScCAE. (A) Growth curve of the uninduced (-tet) and induced (+tet) TbCAE RNAi cell lines (same curve and Northern blot as in Fig. 1A). **(B)** Growth curve of the TbCAE RNAi cell line complemented by expression of the triple c-myc tagged cytosolic/nuclear isoform of yeast ScCAE. Insets: loading control showing ethidium bromide stained (EtBr) rRNA region; Northern blots (NB) of total RNA from uninduced and induced cells probed for TbCAE mRNA and in right panel immunoblots demonstrating inducible expression of ScCAE-myc. **(C)** Left panel: Northern blots (NB) of total cellular RNA isolated from uninduced (-tet) and induced (+tet) TbCAE RNAi cell line probed for tRNA^{Cys} using specific oligonucleotide hybridization. **(D)** Right panel: as (c) but isolated mitochondrial RNA of the TbCAE RNAi cell line complemented with ScCAE-myc was analysed.